

Rituximab target-mediated elimination is influenced by both baseline antigenic burden and *FCGR3A*-158V/F polymorphism in chronic lymphocytic leukemia

Mira Tout¹, Anne-Laure Gagez², Stéphane Leprêtre³, Nicolas Azzopardi¹, Caroline Dartigeas⁴, Béatrice Mahé⁵, Emmanuelle Ferrand⁶, Hervé Maisonneuve⁷, Thérèse Aurrant⁸, Gilles Paintaud^{1,9}, Guillaume Cartron^{2,10}, David Ternant^{1,9}

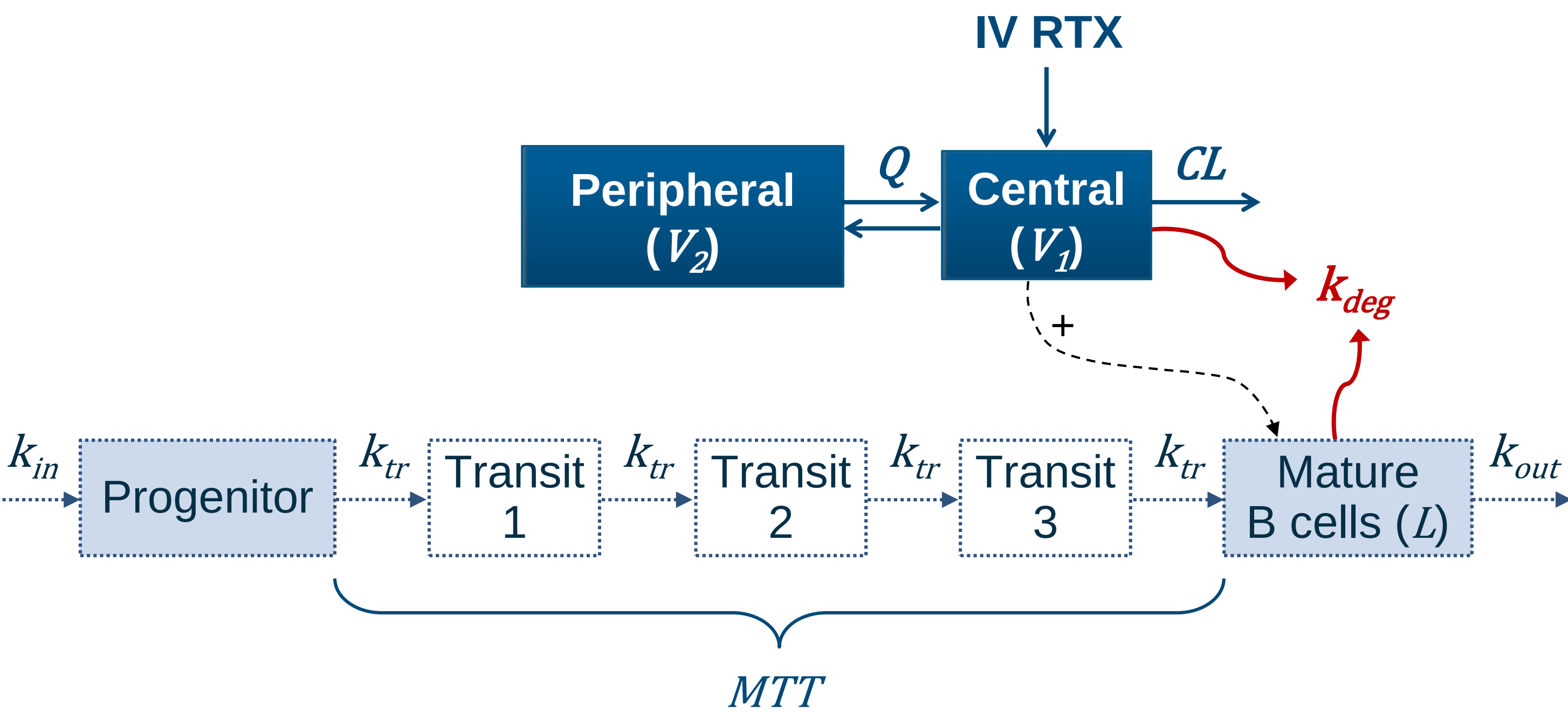
¹CNRS UMR 7292, Université François-Rabelais, Tours, France, ²CNRS UMR 5235, Université de Montpellier, Montpellier, France, ³Hematology, Henri Becquerel Center, Rouen, France, ⁴Hématologie et thérapie cellulaire, CHRU Tours, Tours, France, ⁵Hematology, CHRU Nantes, Nantes, France, ⁶Hematology, CHRU Dijon, Dijon, France, ⁷Hematology, CHD La Roche sur Yon, La Roche Sur Yon, France, ⁸Hematology, Institut Paoli Calmette, Marseille, France, ⁹Laboratoire de Pharmacologie-Toxicologie, CHRU Tours, Tours, France, ¹⁰Hematology, CHRU Montpellier, Montpellier France

Background

The anti-CD20 monoclonal antibody rituximab (RTX) is indicated as first-line treatment in chronic lymphocytic leukemia (CLL). Rituximab exhibits time-dependent clearance in CLL [1], possibly because of treatment-related decrease in B-cell antigenic burden over time. The aim of this study was to quantify the effects of CD20 antigenic mass and FcγRIIIA gene polymorphism on rituximab PK in CLL patients.

Patients and methods

- 118 CLL patients from the CLL 2010 FMP phase II study (1943 RTX serum concentrations)
- Dosing regimens:
 - Standard R-FC arm: *Cycle 1: IV RTX 375 mg/m² + FC (fludarabine, cyclophosphamide); *Cycles 2-6: IV RTX 500 mg/m² + FC; Q 28 days
 - Dense R-FC arm: *Prephase: IV RTX (500 mg on day 0; 2000 mg on days 1, 8, 15); *Cycles 1-6: IV RTX 500 mg/m² + FC Q 28 days
- Population modelling using Monolix® 4.3.3
- Semi-mechanistic model including two compartments linked to latent target-antigen turnover
- Evaluated covariates: gender, age, body size, treatment arm, baseline CD20 antigen count in circulation (CD20_{cir}) and lymph nodes (CD20_{LN}), Binet stage, gene polymorphisms (*FCGR3A*-158V/F, *FCGR2A*-131H/R)



$$\begin{aligned} \frac{dA_1}{dt} &= -(CL + Q) \times \frac{A_1}{V_1} + Q \times \frac{A_2}{V_2} - k_{deg} \times A_1 \times L \\ \frac{dA_2}{dt} &= Q \times \left(\frac{A_1}{V_1} - \frac{A_2}{V_2} \right) \\ \frac{dProgenitor}{dt} &= k_{in} - k_{tr} \times Progenitor, \quad Progenitor(0) = L_{ini} \\ \frac{dTransit1}{dt} &= k_{tr} \times Progenitor - k_{tr} \times Transit1 \\ \frac{dTransit2}{dt} &= k_{tr} \times Transit1 - k_{tr} \times Transit2 \\ \frac{dTransit3}{dt} &= k_{tr} \times Transit2 - k_{tr} \times Transit3, \quad Transit1,2,3(0) = L_{ini} \\ \frac{dL}{dt} &= k_{tr} \times Transit3 - k_{out} \times L - k_{deg} \times A_1 \times L, \quad L(0) = L_{ini} \\ k_{tr} &= \frac{n+1}{MTT}, \quad L_{ini} = \frac{k_{in}}{k_{out}} \end{aligned}$$

Results

Table 1: Parameter estimates

Parameter (unit)	Estimate	RSE (%)	p
Fixed effects			
CL (L/d)	0.137	4	
Dense R-FC arm on CL	0.197	29	6.7x10 ⁻⁴
V ₁ (L)	3.08	2	
BSA on V ₁	0.904	21	2.5x10 ⁻⁶
Q (L/d)	0.31	3	
V ₂ (L)	2.54	6	
Dense R-FC arm on V ₂	0.52	16	2.9x10 ⁻¹⁰
BSA on V ₂	1.31	30	10 ⁻³
MTT (d)	57.6	5	
k _{in} (nmol/d)	0.22	6	
k _{out} (d ⁻¹)	17.6x10 ⁻⁵	8	
Binet stage C on k _{out}	-0.392	27	2.1x10 ⁻⁴
k _{deg} (nmol ⁻¹ d ⁻¹)	0.0017	12	
FCGR3A-158VV on k _{deg}	1.47	25	8.6x10 ⁻⁵
CD20 _{cir} on k _{deg}	0.32	25	8.1x10 ⁻⁵
Interindividual and residual variability			
ω _{CL} (%)	29.9	7	
ω _{V1} (%)	18	9	
ω _{V2} (%)	34.7	9	
ω _{kout} (%)	46.3	8	
ω _{kdeg} (%)	90.5	10	
σ _{add} (nmol/L)	8.33	12	
σ _{prop} (%)	20.7	2	

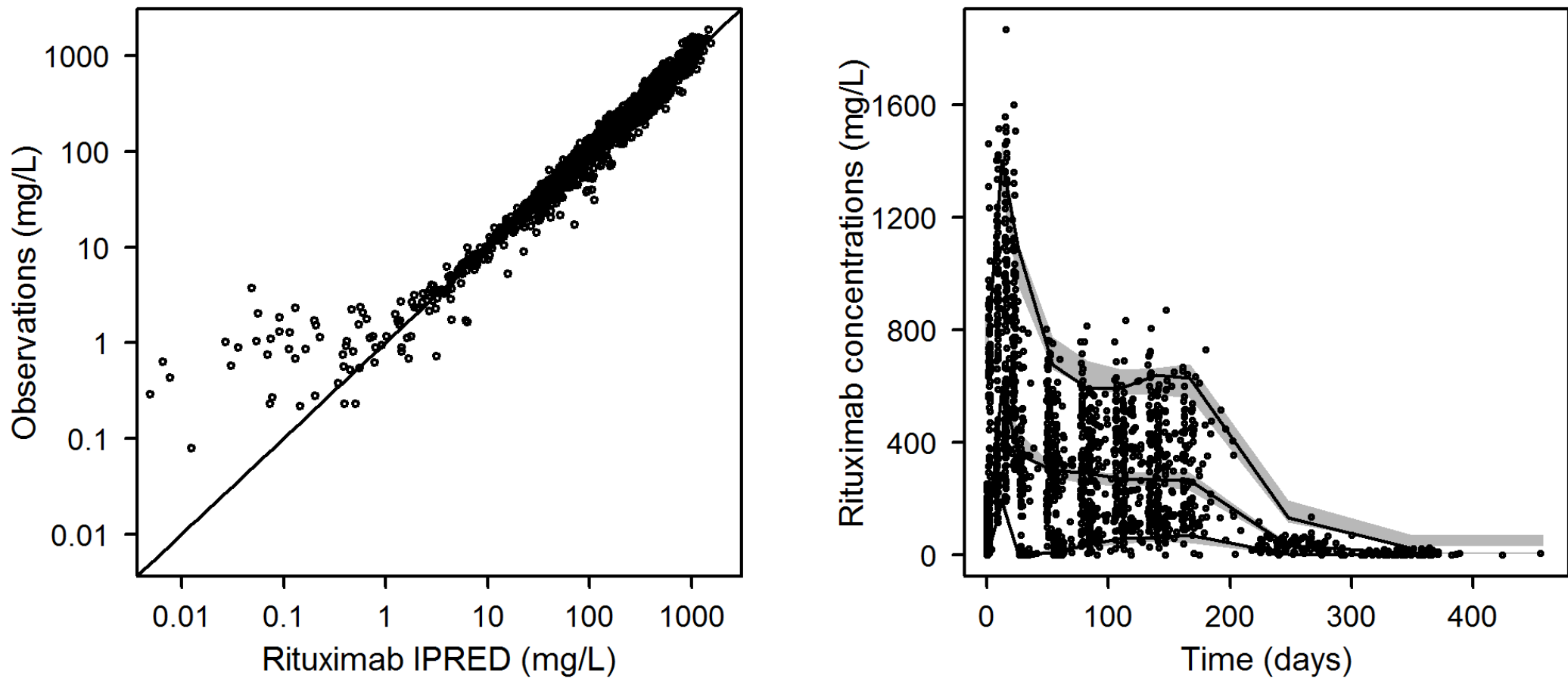
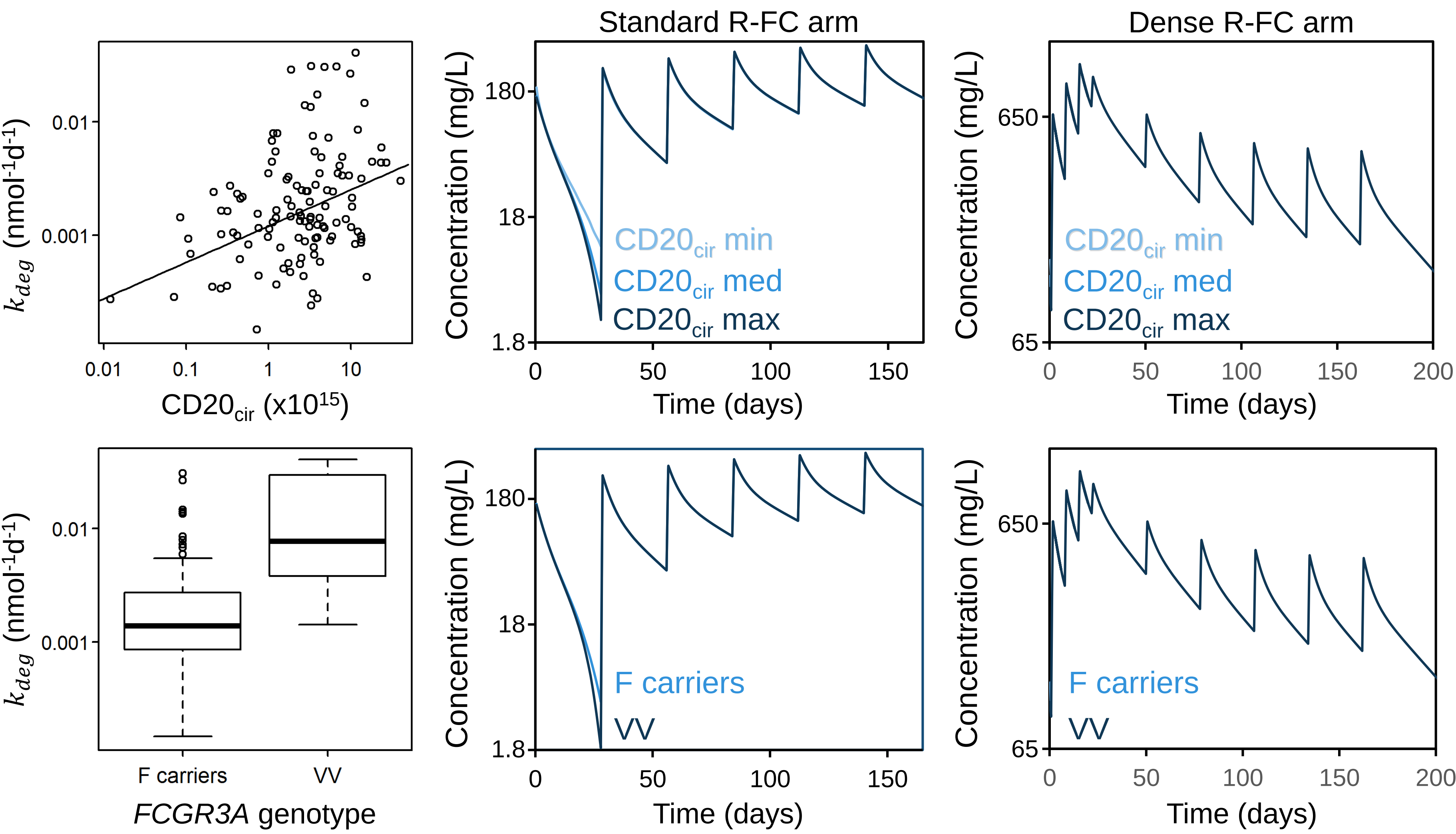


Figure 1: Model diagnostics: Observed vs model-predicted RTX concentrations (left); Visual predictive checks (right)

Model-based simulations



Conclusions

- The developed semi-mechanistic model accurately predicted RTX concentrations
- RTX target-mediated elimination, hence its “consumption”, increased with CD20 target antigen count on circulating B-cells (CD20_{cir}) and in FcγRIIIA-158VV patients
- This influenced RTX concentrations in the first treatment cycle of the standard R-FC arm

References

- [1] Li J. et al. Population pharmacokinetics of rituximab in patients with chronic lymphocytic leukemia. J Clin Pharmacol (2012) 52,1918–26